



DNP[™] Kit High yield DNA Purification Kit

For Research Use Only

SINACLON

Quantity: 50 preps (30 preps for Whole Blood)

Catalog Number: EX6071

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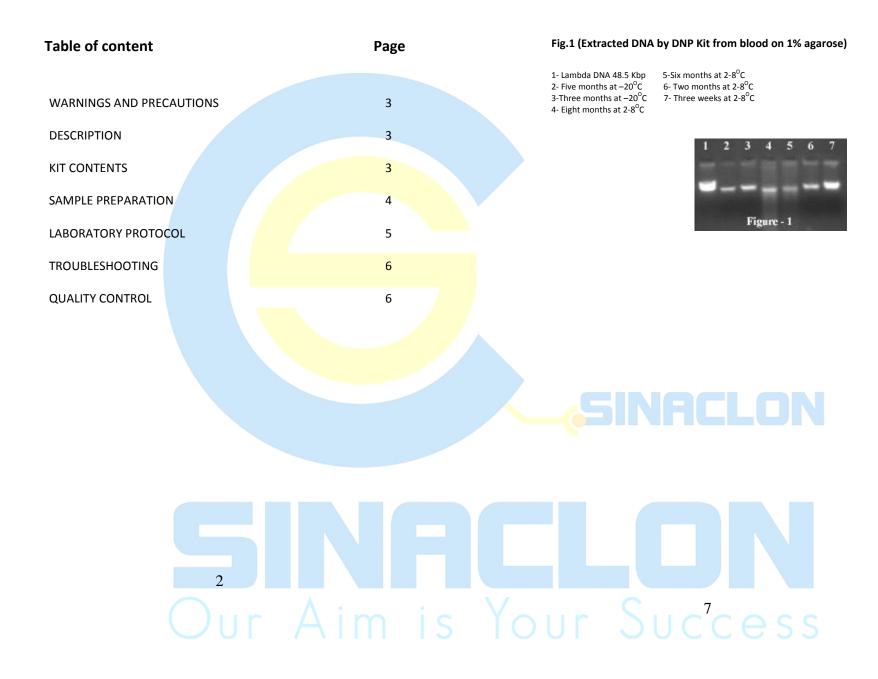
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 Use 1-10μl of DNA solution for each 50μl of PCR mixture. In case of high background PCR product, extracted template DNA may dilute 1/100 and repeat PCR reaction.

TROUBLESHOOTING

Sample	Possible reason	Solution
Low Yield of DNA		
Blood	Sample was frozen and thawed several times. Sample was stored at 2-4°C longer than 2 weeks. Blood clots were present in the sample.	Take new sample.
Cell culture	Too much cells were used for purification, and DNA pellet turned insoluble. Sample was frozen and thawed several times.	Reduce cell quantity twice or more. Take new sample.
Degraded DNA		
Any sample	Inappropriate storage conditions of the sample.	See Sample preparation section.
RNA Contamination Present		
Tissue, bacteria, cell cultures		Add of Ribonuclease A to a final concentration 0.2mg/ml to the sample before step 8) and incubate for 10min at 37°C.
Enzymatic Reactions not Running		
Blood	Heparinized blood was used. Deep Red to Black pellet produced in step 5	Use EDTA or citrate treated blood. Pre-warm Lysis Solution at 37°C and repeat extraction
Any sample	Residual reagents (Wash buffer, salts, etc.) present in prepared DNA, because of inaccurate handling during step 5.	Repeat extraction with once more Wash Buffer tretment.

QUALITY CONTROL

All components of the Kit are successfully tested in the DNA purification and amplification reaction for:

Frozen or fresh whole blood for Thalassemia gene, 50 mg of stomach biopsy for *H. pylori*, one colony of cultured bacteria for RAPD technique, 100 µl of homogenized sputum for MTB and 100µl of positive serum for HBV.



WARNINGS AND PRECAUTIONS

Avoid contact any kit reagents with skin & eyes. Wear gloves before use DNPTM Kit. Contact of Lysis Solution with <u>acids</u> or <u>bleach</u> solution, liberates toxic gas. When handling human samples, follow recommended procedures for biohazardous materials.

DESCRIPTION

Easy to use DNPTM Kit is designed to isolate double-stranded DNA from human or animal sources. The procedure requires 35-55 minutes and does not require phenol extraction nor changing tube during procedure. DNA isolation is based on lysis of the cells and subsequent selective DNA precipitation. Finally, the insoluble DNA is washed and desalted by Wash Buffer. DNA obtained by this method can be used for all molecular biology procedures (PCR, restriction digestion, cloning, Southern blot, DNA sequencing, etc.).

KIT CONTENTS:

Dur Aim is Your Succe³ss

Protease (20 mg /ml) 250 μl

Protease Buffer 5 ml

Lysis Buffer 20 ml

Precipitation Solution (Isopropanol Base) 15 ml

Wash Buffer (Ethanol Base) 2 ×50 ml

Solvent Buffer 2×1250 μl

Note: Before use please prepare RNase A (Cat. No.: MO5411)



-Blood

Whole blood must be collected in EDTA(1mg/ml)- to prevent clotting and DNA degradation. DNA extracted from heparinized blood cannot be used for PCR.

Typically 100 μ l of fresh blood is used for DNA isolation with the yield of 1.0-5.0 μ g . Add 100 μ l of **Protease Buffer** to 100 μ l of fresh blood in a 1.5ml microcentrifuge tube and then add 5 μ l of **protease**, place in 55°C for 30 min. If the blood is to be stored for later use it can be left at 2-4°C for (no longer than) 2 weeks. For long-term storage the samples should be aliquoted in 100 μ l portions and kept at -20°C.

-Sera

Add 5 μ l of **Protease** to 100 μ l of serum or plasma in 1.5 ml microcentrifuge tube, vortex and placed in 72°C for 10 min and then follow the laboratory protocol. Usage of fresh sera samples is recommended. If it is impossible, samples should be frozen in aliquots of 100 μ l and stored at -20°C.

-Homogenized sputum for detection of M. tuberculosis

Add 100 μ l of **Protease Buffer** to 100 μ l of homogenized sputum sample Placed in 1.5 ml microfuge tube and then add 5 μ l **Protease**, mix and then place at 55°C for 30-60 min and then place at 95°C water bath for 20 min. and follow the protocol.

-CSF

Centrifuge 1- 1.5 ml of CSF specimen at 10.000 g for 5 min. and discard supernatant. Add 100 μl **Protease Buffer**, mix and then add 5 μl of **Protease** place at 55°C for 30 min and follow the protocol.

-Cell Culture

Collect cells by centrifugation and resuspende in 100µl of **Protease Buffer**. Add 5 µl of **Protease** to resuspende cells, mix and then place in 55°C for 30-60 min and then follow the laboratory protocol. For the efficient DNA separation from cell components is not recommended to use more cells than 4.0-6.0×10⁶. Usage of thawed cells is not recommended.

-Tissues

Add 100 μ l of **Protease Buffer** to 25-50mg of mammalian tissue (either fresh or frozen) placed in a 1.5ml microcentrifuge tube and then add 5 μ l of **protease**, place in 55°C for 1 to 3 hr (up to degrade) and then follow the laboratory protocol. Usage of thawed samples is not recommended.

-Bacterial Cultures (for PCR application)

Bacterial culture should be centrifuged for 10min at 7500g. Collect 2×10^9 or 10-20mg of bacterial culture (either fresh or frozen) in a 1.5ml microcentrifuge tube and suspend it in 100μ l of **Protease Buffer**. Add 5 μ l of **Protease** to suspension, mix and then place at 55°C for 30 min and follow the laboratory protocol. Usage of thawed cells is not recommended.

-Buccal or Vaginal Swab for detection of bacterial genome

Air-dry the swab for at least 2 hr after collection. Place air-dry swab in 1.5ml microfuge tube, contains 200 μ l sterile distillated water and incubate at 56°C water bath for 30 min. Press the stem end of tube towards the swab two to three times and then remove swab and place sample at 95°C water bath for 10 min then follow the protocol.

Vortexing during incubation time may increase the DNA yield (every 5 minutes for 5 sec)



LABORATORY PROTOCOL

- -Pre-warm kit to room temperature before use.
 -Pre-warm Lysis Solution by placing in 37°C for 10 min and softly shake.
- 2. Mix 100µl of sample with 400µl of Lysis Solution (700µl for whole blood) and vortex 15-20 sec. The sample should be completely homogenous suspension at this step. Any aggregation, clot or insoluble materials could be degraded by softly pipetting or removed.
- 3. Add 300μl of Precipitation Solution (For whole blood add 500 μl), mix by vortexing 5 seconds, then centrifuge 12,000 g for 10min.
- 4. Decant by gently inverting of tube and placing the tube on tissue paper for 2-3 sec. down ward. Care for avoid of cross-contamination between different samples.
- 5. Add **1 ml Wash Buffer** to pellet, mix by <u>3-5 seconds vortexing</u> and centrifuge at 12,000 g for 5min, then decant (for whole blood and tissue sample repeat this step once more).
- 6. Pour off the **Wash Buffer** completely and dry pellet at 65°C for 5 min. (up to dry).
- 7. Suspend pellet in **50μl** of **Solvent Buffer** (for **serum** or **plasma** sample, suspend pellet in **30μl**) by gentle shaking and placing at 65°C for 5 min. Wash the wall of tube for mixing of any residual pellet by softly pippeting.
- 8. Precipitate unsolved materials by centrifuge, 30 sec at 12,000 g, supernatant contains purified DNA. Measure DNA concentration spectrophotometrically or visually after electrophoresis in fresh 1% agarose gel. (fig.1)

Note:

- For all sample If RNA-free genomic DNA is required, add <u>Ribonuclease A</u> (final concentration 0.2mg/ml), to the sample during protease incubation.
- Transcriptionally active tissue and bacterial culture contain high levels of RNA, which can
 copurify with genomic DNA. RNA may inhibit some downstream enzymatic reactions, but
 not the PCR itself. If RNA-free genomic DNA is required, add <u>Ribonuclease A</u> (final
 concentration 0.2mg/ml), to the sample during protease incubation.
- For whole blood, DNA yield depends on quantity of leukocyte cells and storage duration and condition of sample.
- Since DNA quantity is too small, Viral DNA from sera samples is invisible in agarose gel and only host nucleic acids from lysed leucocytes may be monitored